

Chestnut Chat GWAS 101

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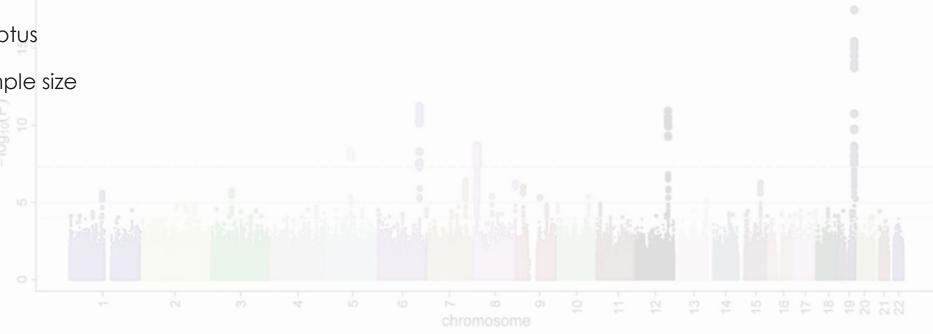


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GWAS 101

- 1. Overview of GWAS
- 2. What kind of traits can be analyzed?
- 3. Phenotypes and Genotypes
- 4. The three steps of GWAS
- 5. A case study in Eucalyptus
- 6. The importance of sample size
- 7. Population Structure
- 8. Interpreting results





GWAS according to the NIH

A genome-wide association study (GWAS) is an approach used in genetics research to associate specific genetic variations with particular diseases.

The method involves scanning the genomes from many different people and looking for genetic markers that can be used to predict the presence of a disease. Once such genetic markers are identified, they can be used to understand how genes contribute to the disease and develop better prevention and treatment strategies.

GWAS according to me

Phenotype = Genotype + Environment



GWAS for pathogen resistance in trees

Poplar

Association mapping, transcriptomics, and transient expression identify candidate genes mediating plant-pathogen interactions in a tree

Wellington Muchero^a, Kelsey L. Sondreli^b, Jin-Gui Chen^a, Breeanna R. Urbanowicz^c, Jin Zhang^a, Vasanth Singan^d, Yongil Yang^a, Robert S. Brueggeman^e, Juan Franco-Coronado^e, Nivi Abraham^e, Jeong-Yeh Yang^c, Kelley W. Moremen^c, Alexandra J. Weisberg^b, Jeff H. Chang^b, Erika Lindquist^d, Kerrie Barry^d, Priya Ranjan^a, Sara Jawdy^a, Jeremy Schmutz^{d,f}, Gerald A. Tuskan^{a,d}, and Jared M. LeBoldus^{b,e,g,1}

^aBiosciences Division, Oak Ridge National Laboratory, Oak Ridge, TN 37831; ^bDepartment of Botany and Plant Pathology, Oregon State University, Corvallis, OR 97331; ^CComplex Carbohydrate Research Center, University of Georgia, Athens, GA 30602; ^dJoint Genome Institute, US Department of Energy, Walnut Creek, CA 94598; ^eDepartment of Plant Pathology, North Dakota State University, Fargo, ND 58102; ^fHudsonAlpha Institute for Biotechnology, Huntsville, AL 35806; and ^gForest Engineering, Resources, and Management, Oregon State University, Corvallis, OR 97331

Ash

ARTICLES https://doi.org/10.1038/s41559-019-1036-6 ecology & evolution

Genomic basis of European ash tree resistance to ash dieback fungus

Jonathan J. Stocks^{1,2}, Carey L. Metheringham^{(3)1,2}, William J. Plumb^{1,2,3}, Steve J. Lee⁴, Laura J. Kelly^{(3)1,2}, Richard A. Nichols¹ and Richard J. A. Buggs^{(3)1,2}*

Eucalyptus

A Genome-Wide Association Study for Resistance to the Insect Pest *Leptocybe invasa* in *Eucalyptus grandis* Reveals Genomic Regions and Positional Candidate Defense Genes

Lorraine Mhoswa¹, Marja M. O'Neill¹, Makobatjatji M. Mphahlele^{1,2}, Caryn N. Oates¹, Kitt G. Payn³, Bernard Slippers¹, Alexander A. Myburg¹ and Sanushka Naidoo ¹/_{*}

Beech

RESEARCH ARTICLE

Open Access

Genome-wide association study identifies a major gene for beech bark disease resistance in American beech (*Fagus grandifolia* Ehrh.)

Irina Ćalić¹, Jennifer Koch², David Carey², Charles Addo-Quaye^{3,4}, John E. Carlson⁵ and David B. Neale^{1*}

Eucalyptus

New Phytologist



Regional heritability mapping and genome-wide association identify loci for complex growth, wood and disease resistance traits in *Eucalyptus*

Rafael Tassinari Resende¹, Marcos Deon Vilela Resende^{1,2}, Fabyano Fonseca Silva³, Camila Ferreira Azevedo¹, Elizabete Keiko Takahashi⁴, Orzenil Bonfim Silva-Junior^{5,6} and Dario Grattapaglia^{5,6}



GWAS for wood properties

| Phenotype | Species | Population | Sample size | No. of markers | Method | Reference |
|---|--|-------------------------------|-------------|---|---|----------------------------|
| Growth and wood properties | Eucalyptus globulus | Families and bulk collections | 303 | 7,680 [Diversity Array Technology markers (DArT)] | General linear model (GLM) and unified mixed model (UMM) | Cappa et al., 2013 |
| Wood density, stiffness, microfibril angle, and ring width | Picea glauca | Open-pollinated families | 1694 | 7434 (SNPs) | Mixed linear model (MLM) | Lamara et al., 2016 |
| 16 wood chemistry/ ultrastructure traits | Populus trichocarpa | Unrelated individuals | 334 | 29,233 (SNPs) | GLM | Porth et al., 2013 |
| Lignin percentage, Lignin S:G ratio, 5-carbon sugars, and 6-carbon sugars | Populus deltoides | Unrelated individuals | 391 | 334,679 (consensus SNPs), 185,526 (Common SNPs), 76,804 (functional SNPs) | Single-variant and multiple-variant associations on GLM | Fahrenkrog et al., 2017 |
| Basic wood density (BWD), bleached oulp, pulp yield (SPY), and pulp bleaching content | Eucalyptus grandis × Eucalyptus urophylla | Hybrid breeding population | 768 | 24 806 (SNPs) | GWAS and regional heritability mapping | Resende et al., 2017 |
| 17 wood-quality traits | Norway spruce | Mother trees | 517 | 178101 (SNPs) | Multilocus LASSO penalized regression | Baison et al., 2018 |
| Seven wood properties | Populus tomentosa | Unrelated individuals | 435 | 5,482 (InDels) | MLM and Kempthorne model | Gong et al., 2017 |

Diversity Array Technology (DArT) markers.

Du, Qingzhang, et al. "Genome-wide association studies to improve wood properties: challenges and prospects." *Frontiers in Plant Science* 9 (2018): 1912.



You can GWAS almost anything you can measure

Typical GWAS traits in trees

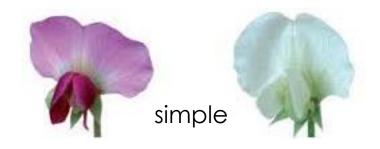
- Growth: Height, DBH, crown architecture, branching, leaf shape
- Wood properties: lignin, sugars, microfibril angle
- Metabolic: primary or secondary metabolite abundances
- Pathogens: Fungal/bacterial abundances
- Resistance: inoculation outcomes
- Adaptive: flowering time, leaf senescence
- Sustainability: water use efficiency, nitrogen use
- Gene Expression: Gene Transcript (RNAseq) abundances

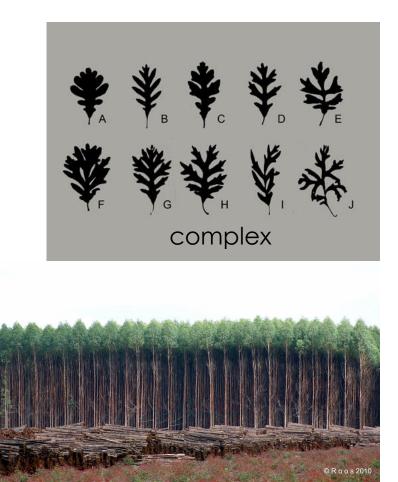


Phenotypic variation

- Without variation there is little to explore
- Variation is the source material
- Is the phenotypic variation due to:
 - Genetic variation?
 - Environmental variation?
 - Both?

Phenotype = Genotype + Environment



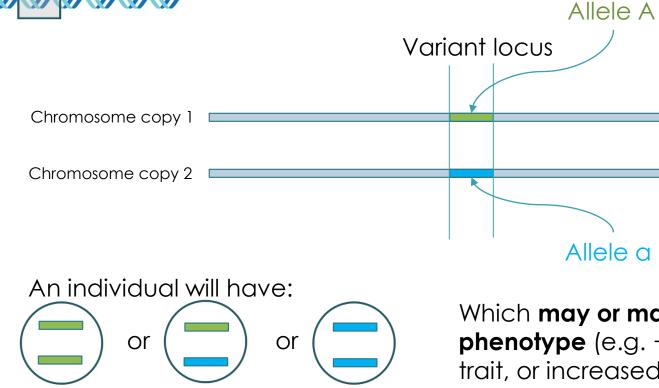






genetic variation

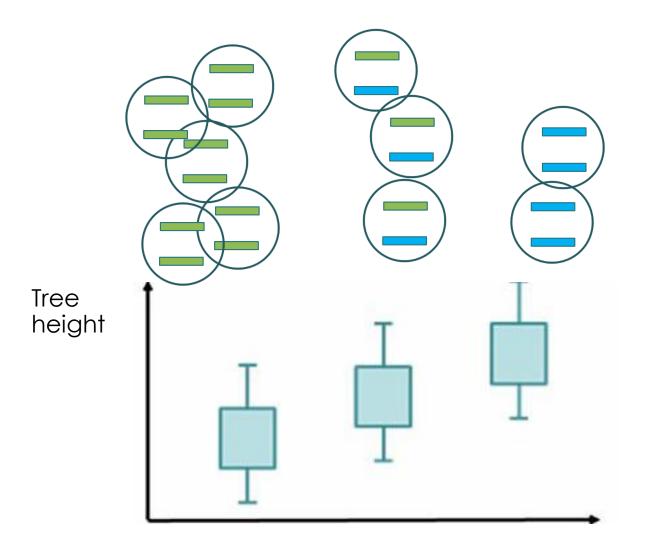
Diploid "chromosome"



Which **may or may not affect their phenotype** (e.g. +/- for quantitative trait, or increased resistance to a disease)

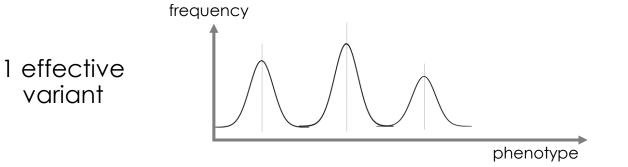


genetic variation in a population





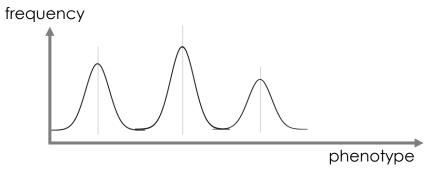
Population Genetic and Phenotypic variation



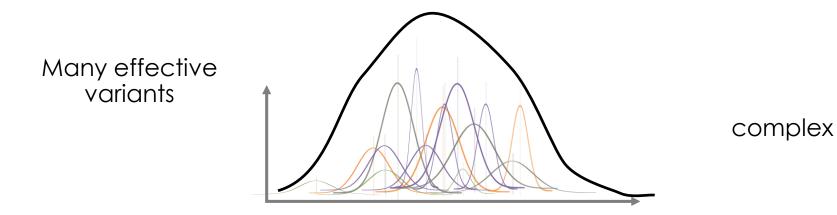
simple



Population Genetic and Phenotypic variation

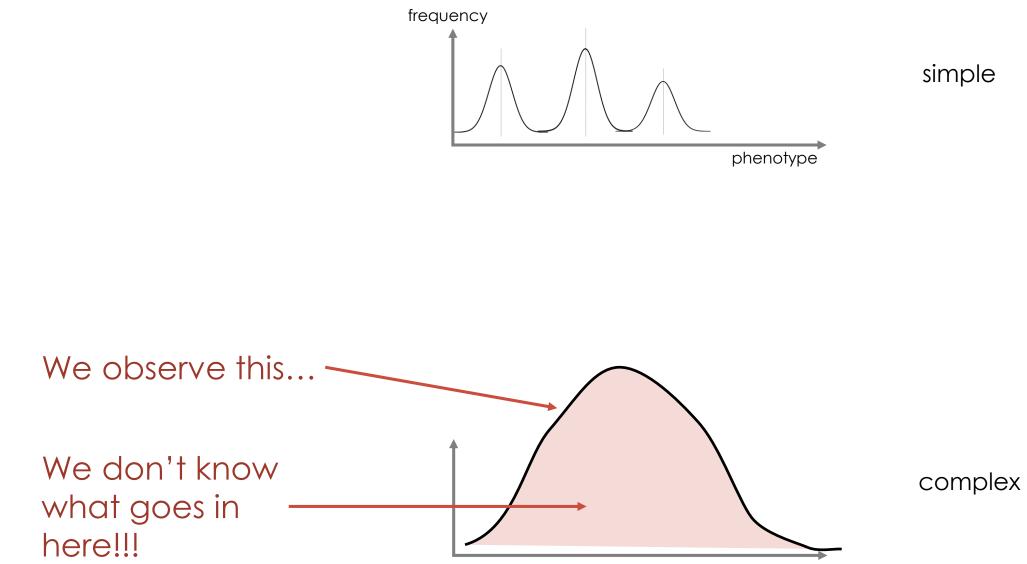


simple





Population Genetic and Phenotypic variation





Genome-wide variation

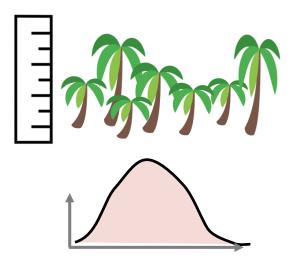


Across the entire genome, there may be millions of variant locations (e.g. SNPs) that have alleles in a population.

Q: Which ones have an effect on a trait ? A: GWAS



How a GWAS works: quantitative trait



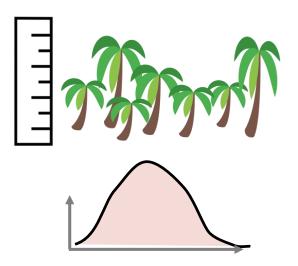


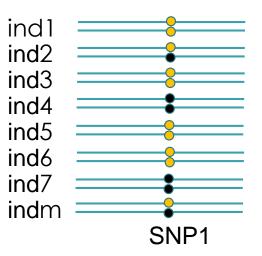
Take a population of mostly <u>un-related</u> individuals. Measure a phenotype that varies (e.g. Height)



How a GWAS works: quantitative trait

2





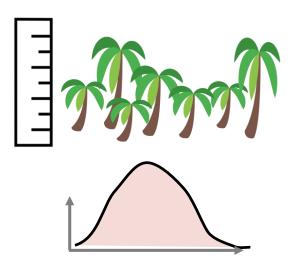


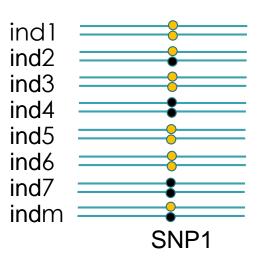
Take a population of mostly <u>un-related</u> individuals. Measure a phenotype that varies (e.g. Height)

- Sequence the DNA of each of them. Find positions in the
- genome where the individuals vary (e.g. SNPs)



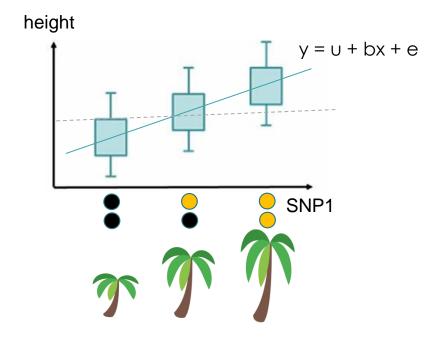
How a GWAS works: quantitative trait







Take a population of mostly <u>un-related</u> individuals. Measure a phenotype that varies (e.g. Height) 2 Sequence the DNA of each of them. Find positions in the genome where the individuals vary (e.g. SNPs)



3 Test each SNP to see if the alleles significantly correlate with the phenotypic variation. E.g. Does having more copies of the • allele equate to a significant linear increase in height?



How a GWAS works: binary trait

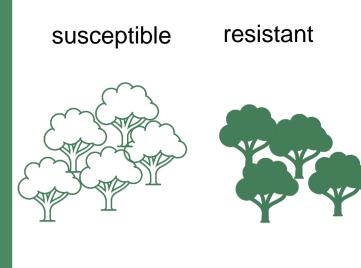
ind1

ind2

ind3

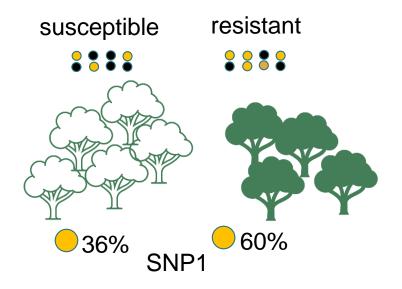
ind4 ind5

ind6 ind7 indm



Take a population of mostly <u>un-related</u> individuals. Split them into two groups (e.g. resistant / susceptible) Sequence the DNA of each of them. Find positions in the genome where the individuals vary (e.g. SNPs)

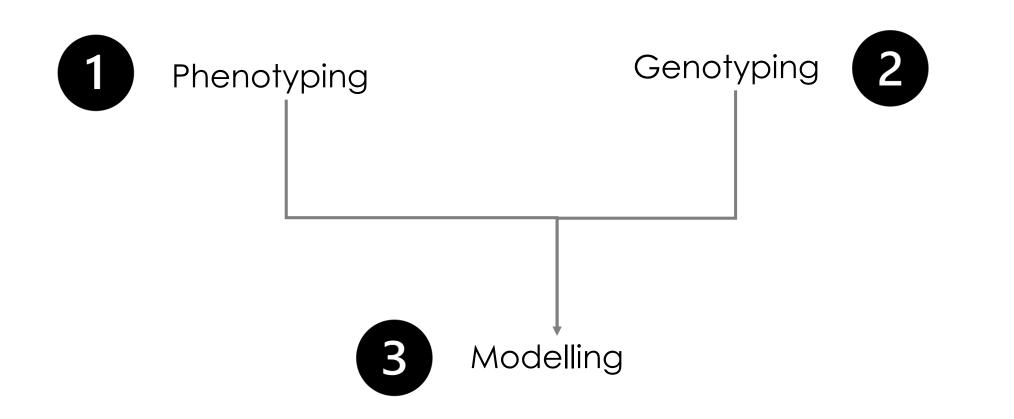
SNP1



Test each SNP to see if its alleles have significantly different frequency in one group compared to the other

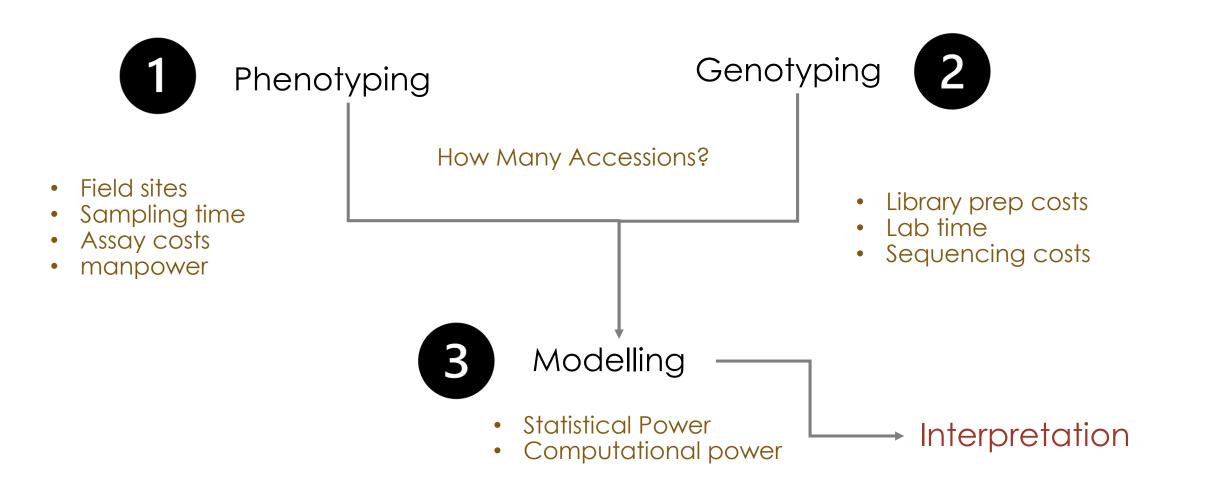


Three main steps for GWAS





Three main steps for GWAS



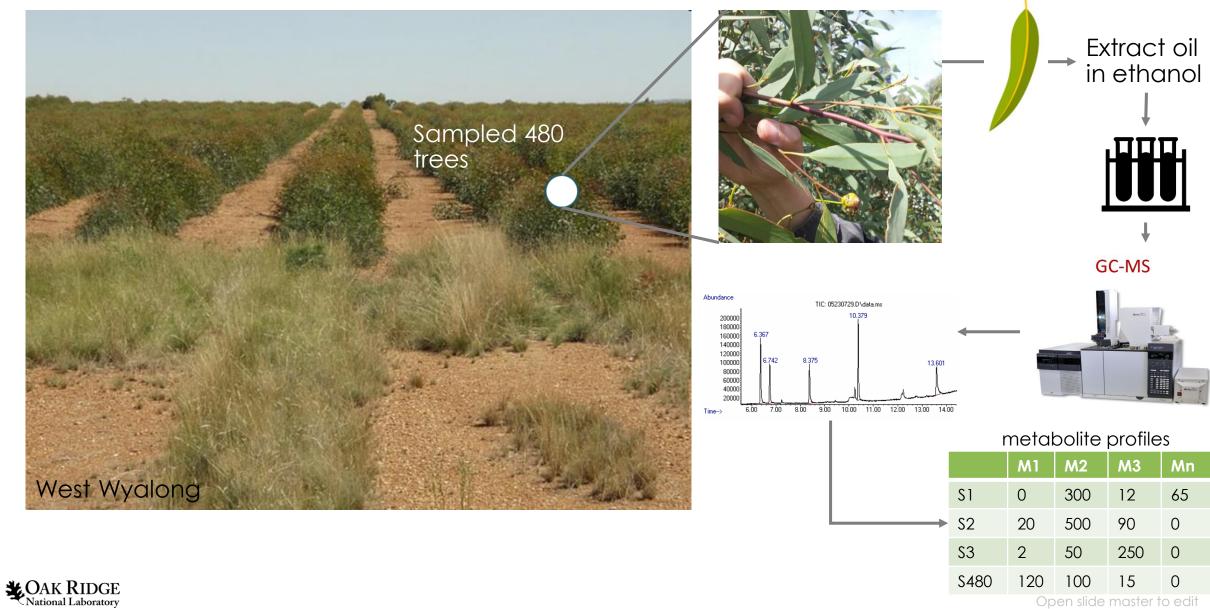


GWAS for oil traits in Eucalyptus polybractea

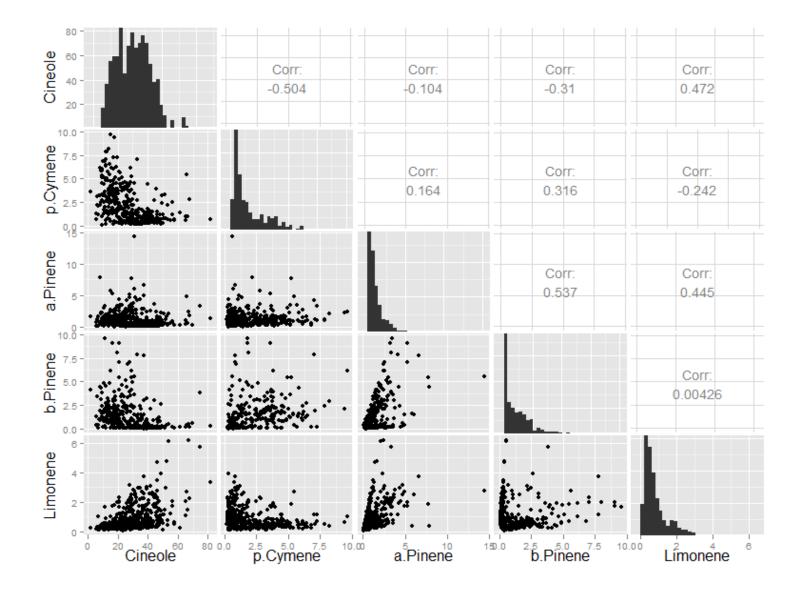














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Phenotyping pitfall – environmental variance





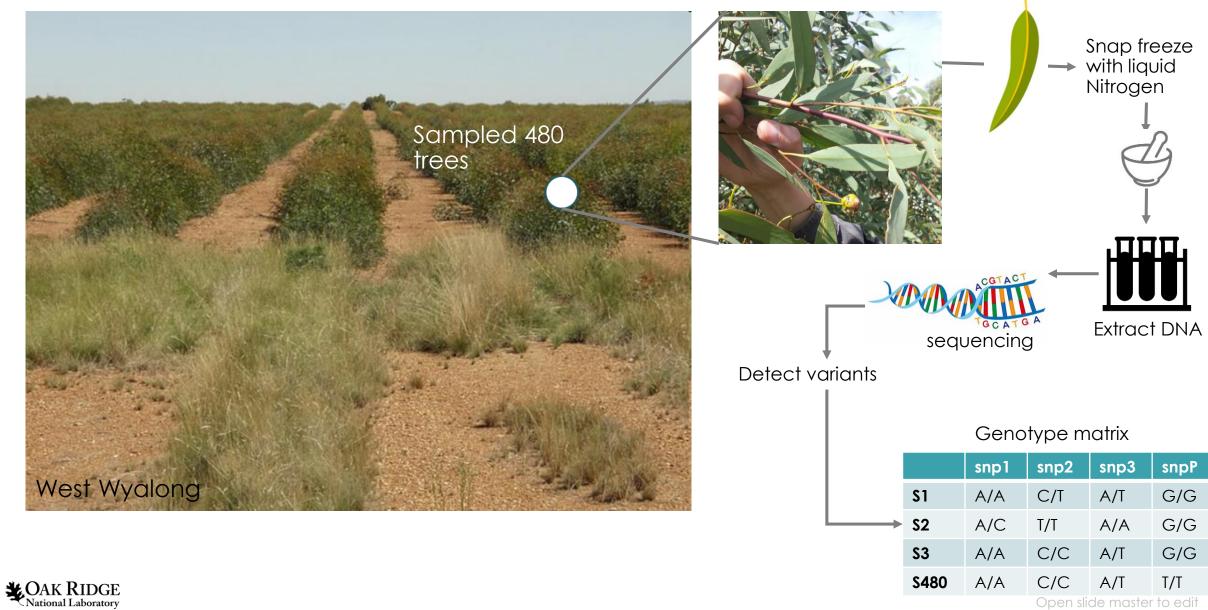


- Minimize environmental variance
- Randomization is necessary to avoid batch effects or environmental trends
- Accuracy is hugely important
- consistency





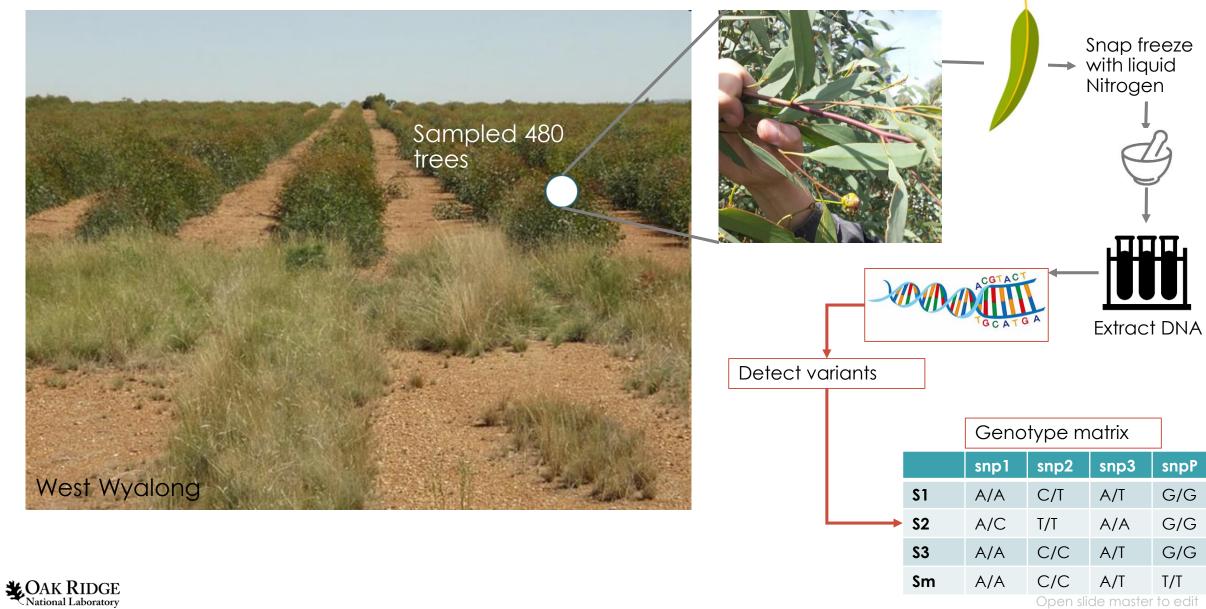
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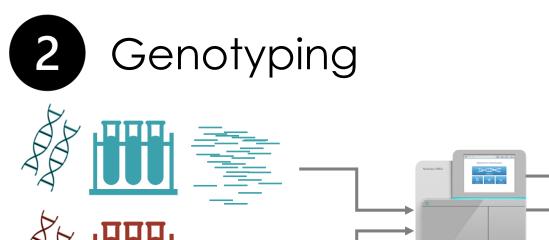
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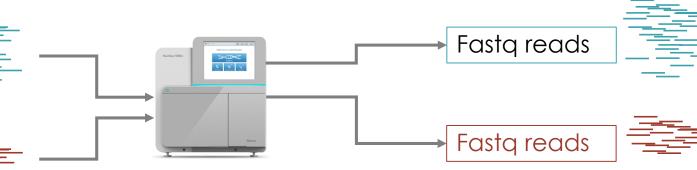


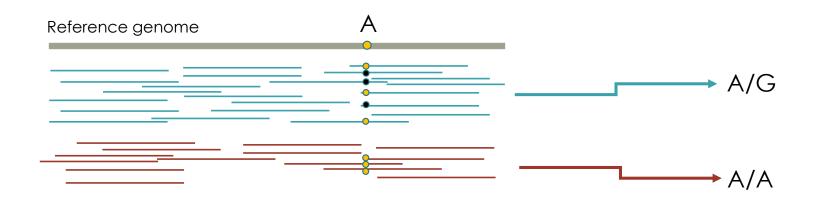
27



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Genotyping pitfall: More accessions or more sequencing depth?

N accessions

2N accessions



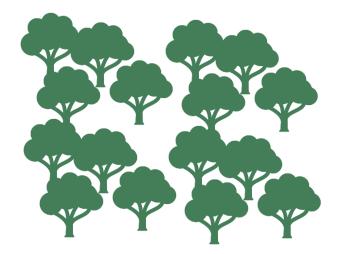
Or...



CAK RIDGE

National Laboratory

- High sequencing depth per sample
- Higher genotype accuracy
- Lower genetic diversity
- Lower statistical power



- low depth per sample
- Lower genotype accuracy
- Higher genetic diversity
- Higher statistical power





metabolite matrix

| | M1 | M2 | M3 | Mn |
|------|-----|-----|-----|----|
| S1 | 0 | 300 | 12 | 65 |
| S2 | 20 | 500 | 90 | 0 |
| S3 | 2 | 50 | 250 | 0 |
| S480 | 120 | 100 | 15 | 0 |

Genotype matrix

| | snp1 | snp2 | snp3 | snpP |
|------------|------|------|------|------|
| S 1 | A/A | C/T | A/T | G/G |
| S2 | A/C | T/T | A/A | G/G |
| S3 | A/A | C/C | A/T | G/G |
| S480 | A/A | C/C | A/T | T/T |
| | Ļ | Ļ | ļ | Ļ |
| | snp1 | snp2 | snp3 | snpP |

| | snp1 | snp2 | snp3 | snpP |
|------------|------|------|------|------|
| S 1 | 0 | 1 | 1 | 0 |
| S2 | 1 | 2 | 0 | 0 |
| S 3 | 0 | 0 | 1 | 0 |
| S480 | 0 | 0 | 1 | 2 |



31



metabolite matrix

| | M1 | M2 | M3 | Mn |
|------|-----|-----|-----|----|
| S1 | 0 | 300 | 12 | 65 |
| S2 | 20 | 500 | 90 | 0 |
| S3 | 2 | 50 | 250 | 0 |
| S480 | 120 | 100 | 15 | 0 |

snp2 snp3 snp1 snpP **S1** A/A C/T A/T G/G **S2** A/C A/A G/G T/T C/C G/G **S3** A/A A/T A/A C/C A/T Sm T/T snp2 snp3 snpP snp1 0 **S1** 0 1 1 **S2** 1 2 0 0 **S**3 0 0 0 1 0 Sm 0 2 1

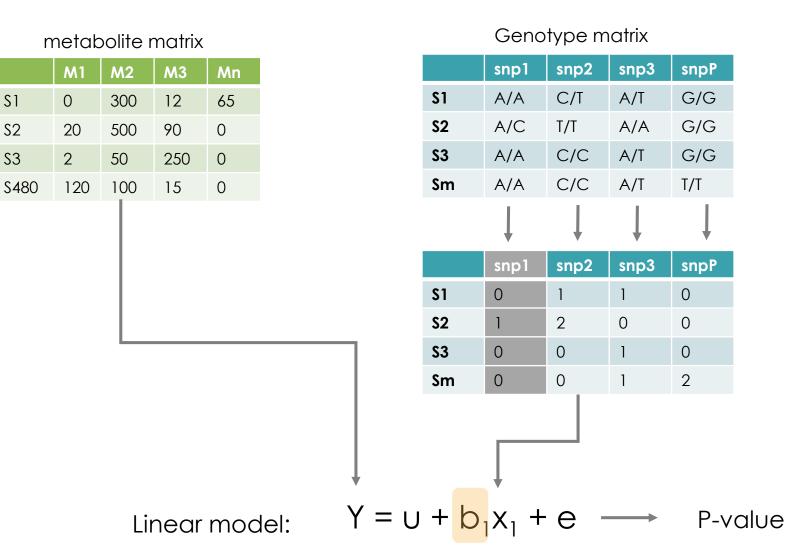
Genotype matrix

Filtering:

- Remove rare SNPs
- Remove SNPs that are likely to be erroneous







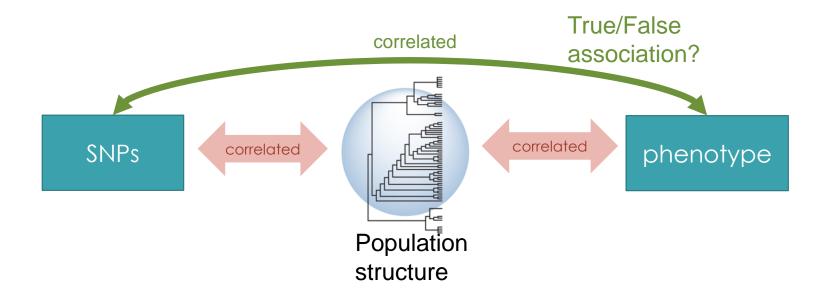


Problem with our model

 $Y = u + b_i x_i + e$

Population structure

 Samples are often genetically related to each other (even if you think they aren't!), which means they are not independent and can cause false positive SNP associations.



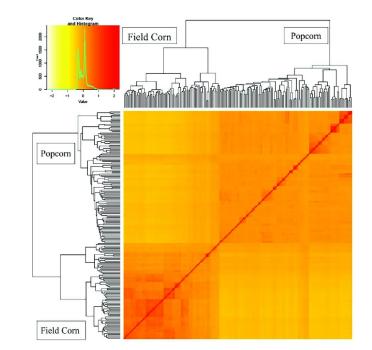


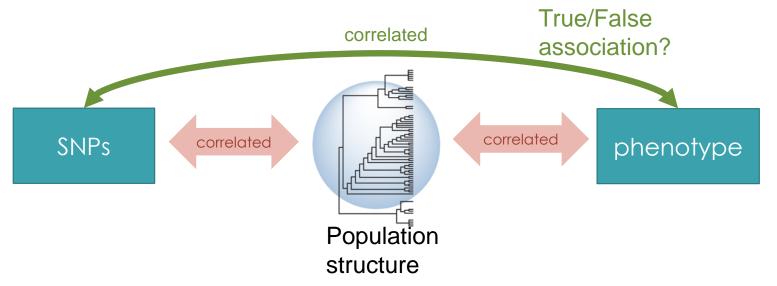
Problem with our model

 $Y = u + b_i x_i + k + e$

Population structure

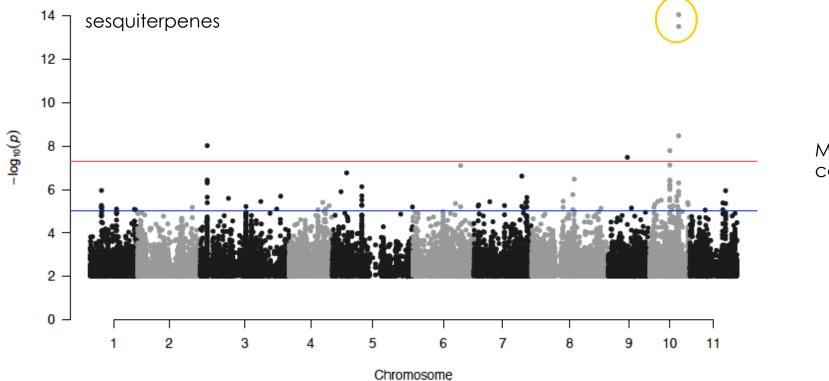
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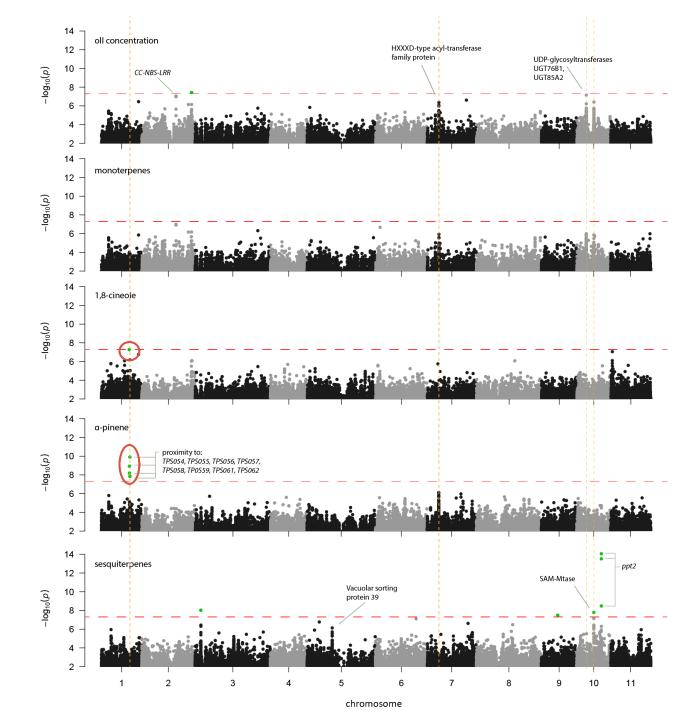
Results: Manhattan plot



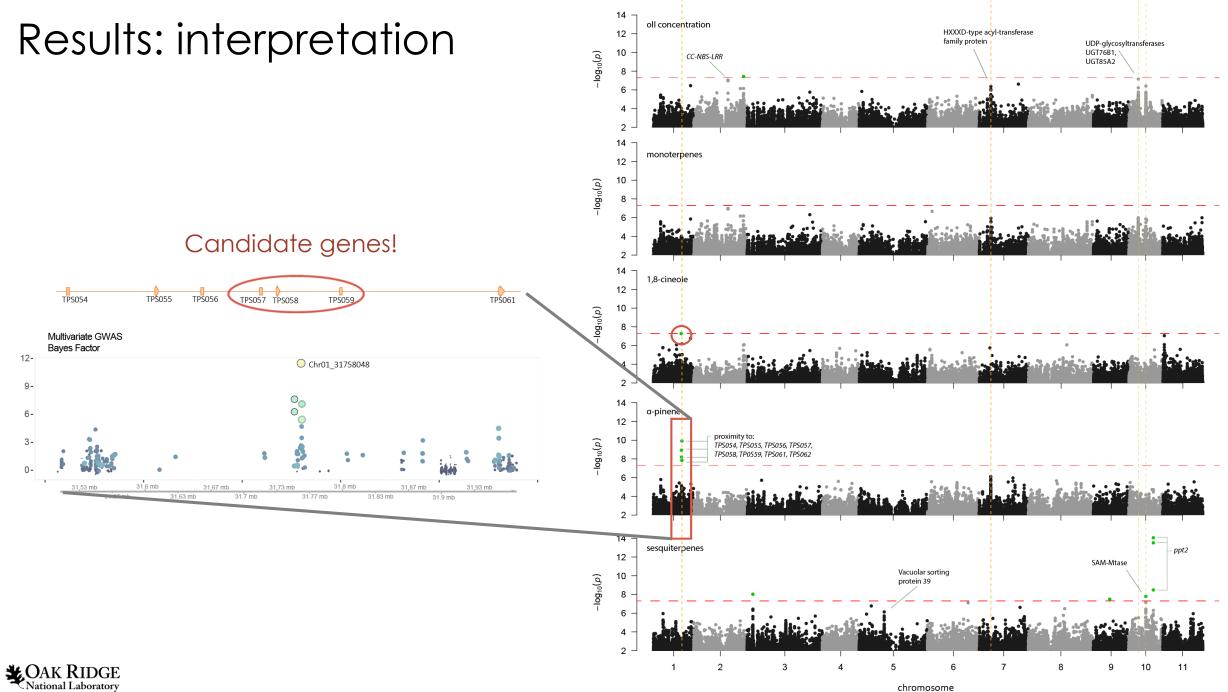
Multiple testing correction threshold



Results





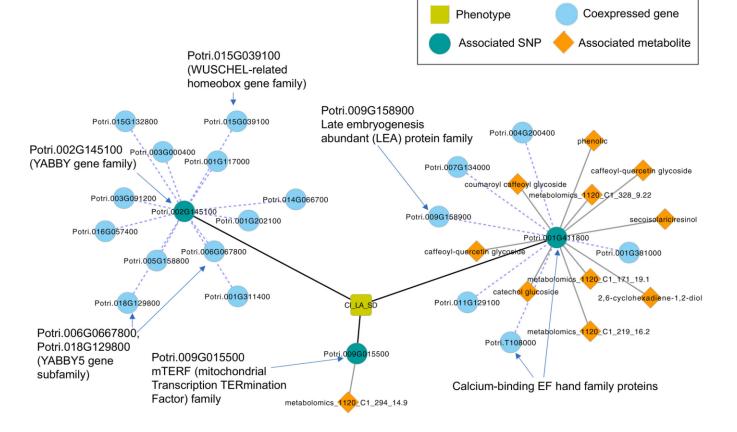


39

chromosome

Next Steps

- Validation !!
- Multi-omics integration
- Try other models
- Build the story





Thank You

- Carsten Kulheim
- William Foley
- Carlos Bustos
- Amanda Padovan
- Daniel Jacobson
- Jerry Tuskan





